

21 **Abstract**

22 This study aimed to prepare fine granules with a diameter less than 200 μm and
23 sustained drug release properties by melt granulation. Triglycerin full behenate
24 (TR-FB) was examined as a new meltable binder (MB) by comparison of its
25 properties with those of glycerin monostearate (GM), a widely used as MB. The
26 effect of milling microcrystalline cellulose (MCC), an excipient for melt granulation,
27 on the granule properties was also investigated. TR-FB was more stable during
28 heating and storage than GM, and produced smaller granules with narrower particle
29 size distribution, larger yield in the 106–200 μm range, uniform roundness and better
30 sustained drug release profile than those prepared with GM. Granules prepared with
31 milled MCC had almost the same physicochemical properties as those produced with
32 intact MCC. However, milled MCC produced granules with a more rigid structure
33 and smaller void space than intact MCC. Consequently, the granules produced with
34 milled MCC showed better sustained drug release behavior than those prepared with
35 intact MCC. We successfully prepared fine granules with sustained drug release
36 properties and diameter of less than 200 μm using TR-FB and milled MCC.

37

38 **Keywords:** High-shear melt granulation; computed tomography; synchrotron X-ray

39 radiation; milling process; internal structure; triglycerin full behenate

40

41 **Abbreviations:** APAP, Acetaminophen; BRH, bromhexine hydrochloride; DSC,

42 differential scanning calorimetry; GM, glycerin monostearate; IS, impeller speeds;

43 MB, meltable binder; MCC, microcrystalline cellulose; PXRD, powder X-ray

44 diffraction; SEM, scanning electron microscopy; TR-FB, triglycerin full behenate;

45 X-ray CT, X-ray computed tomography

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47

48 **1. Introduction**

49 Melt granulation is a granulation method that uses a material with a low
50 melting point as a meltable binder (MB). The granules are formed by agitation or
51 fluidization, and cooling conditions, such as ambient temperature, cause the MB to
52 congeal to give dried granules. This simple process is one advantage melt granulation.
53 In addition, because this method is solvent-free, it does not require a drying step or
54 toxic solvent, so melt granulation is considered an economical and environmentally
55 friendly process (ICH guideline, 1998) that can be applied to water-sensitive drugs.
56 Furthermore, melt granulation can easily provide granules with sustained drug
57 release by using hydrophobic or water-insoluble MBs (Thomsen et al., 1994,
58 Hamdani et al., 2002, Ochoa et al., 2011). To date, numerous functional granules
59 have been prepared by melt granulation; for example, granules with immediate drug
60 release (Perissutti et al., 2003), improved drug solubility (Shah et al., 2013),
61 enhanced stability of a moisture-sensitive drug (Kowalski et al., 2009), and
62 gastroretentivity (Hamdani et al., 2006). Recently, we successfully prepared granules
63 by melt granulation that exhibited pH-dependent drug release (Shiino et al., 2012).

64 Besides their functionality, fine granules with a particle diameter of less than
65 200 μm are desired to improve their taste and texture (Shah, Ket al., 1994). In

66 particular, fine granules that undergo sustained drug release have attracted
67 considerable attention to produce orally disintegrating tablets. However, granules
68 with sustained release properties and diameter of less than 200 μm prepared by melt
69 granulation have not been reported. Schaefer proposed that the smaller the
70 pharmaceutical excipients used, the smaller the granules obtained (Schaefer et al.,
71 1997, Schaefer et al 2004). They used a MB with a mean particle diameter of 40 μm
72 that gave granules with a diameter of approximately 500 μm . This suggests that to
73 prepare fine granules with sustained drug release, hydrophobic or water-insoluble
74 MBs and excipients with a particle size of less than 40 μm should be used. Operating
75 conditions should also be improved to minimize granule size.

76 In the present study, we focus on triglycerin full behenate (TR-FB), which is
77 typically used as a food additive, as a new MB. From a structural perspective, TR-FB
78 has a long carbonyl chain and behenic acid, indicating that it can endow granules
79 with better sustained release properties. In addition, the mean particle diameter of
80 TR-FB is about 4 μm because it is milled by a jet mill (Kamiryo et al., 2002). The
81 maximum acceptable daily intake of TR-FB is 25 mg/kg body weight/day, so it has
82 similar safety to sucrose fatty acid ester, which is often used as a pharmaceutical

83 excipient (FAO/WHO, 1989). Therefore, TR-FB might be suitable for preparing
84 sustained-release fine granules.

85 We also investigated microcrystalline cellulose (MCC) as a pharmaceutical
86 excipient. In general, lactose, starch and calcium hydrogen phosphate have been used
87 as excipients for melt granulation; however, because these excipients are hydrophilic
88 or have large particle size even after milling, they are not considered suitable
89 excipients to prepare fine granules. In contrast, MCC is insoluble in water, easily
90 downsized by milling and has recently been used in melt granulation (Kukec et al.,
91 2012), indicating that it may be a suitable excipient. However, how downsized MCC
92 affects the physicochemical properties of fine granules prepared by melt granulation
93 has not been examined.

94 With the aim of preparing fine granules with sustained drug release, our two
95 main objectives were as follows: the first was to investigate whether TR-FB acts as
96 an MB by comparison with glycerin monostearate (GM), which is commonly used as
97 a hydrophobic MB, and the second was to determine the effects of milling MCC on
98 melt granulation by comparison of the granules prepared with intact and milled MCC.
99 Differential scanning calorimetry (DSC) and powder X- ray diffraction (PXRD) were
100 performed to determine the stability of TR-FB and GM after heat treatment. Next,

101 the physicochemical properties, including particle size distribution, roundness and
102 drug release profile, of granules prepared with either GM or TR-FB were examined.
103 The particle size distribution, roundness, morphology, internal structure, porosity and
104 drug release profile of granules prepared with either intact MCC or MCC milled at
105 different impeller speeds (IS) were then studied.

106

107

108 **2. Materials and methods**

109 **2.1. Materials**

110 Acetaminophen (APAP) and bromhexine hydrochloride (BRH), as model
111 drugs, were kindly provided by Iwaki Pharmaceutical Co., Ltd. (Shizuoka, Japan)
112 and purchased from Shiratori Pharmaceutics Inc. (Chiba, Japan), respectively. MCC
113 PH-102 was kindly provided by Asahi Kasei Chemicals Co., Ltd. (Tokyo, Japan).
114 GM was purchased from Taiyo Chemical Industry Co., Ltd. (Tokyo, Japan). TR-FB
115 was kindly provided by Riken Vitamin Co., Ltd. (Tokyo, Japan). Talc was purchased
116 from Gotoku Chemical Co., Ltd. (Tokyo, Japan).

117

118 **2.2. DSC**

119 Melting points of the MBs were determined by DSC (EXSTAR DSC 7020,
120 SII Nano Technology Inc, Chiba, Japan). Samples (about 10 mg) were sealed in a
121 40-mL aluminum pan. The samples were analyzed during heating from 25 to 200 °C
122 (first run), cooling to 25 °C, and reheating from 25 to 200 °C (second run) at a
123 heating rate of 5 °C/min under a nitrogen atmosphere (40 mL/min).

124

125 **2.3. PXRD**

126 The MBs were analyzed by PXRD (Rigaku Rotaflex RU-200B, Rigaku
127 Corp, Tokyo, Japan) under the following operating conditions: target: Cu; voltage: 30
128 kV, current: 15 mA; scanning speed: 4 °/min; 2 θ range: 3–40°.

129

130 ***2.4. Storage stability of MB particles***

131 GM and TR-FB particles were prepared by the spray congealing method
132 reported in our previous paper (Nitanai et al., 2012) and then stored at 24 °C/25%
133 RH for 6 weeks. The surface morphology of MB particles before and after storage
134 was observed by scanning electron microscopy (SEM; JSM-5310LV, JEOL Ltd.,
135 Tokyo, Japan). The samples were placed on double-sided adhesive tape and sputter
136 coated with platinum under vacuum prior to imaging.

137

138 ***2.5. High-shear melt granulation***

139 ***2.5.1. Granules containing APAP***

140 Size reduction of APAP and MCC was carried out using a sample mill
141 (TI-300, Cosmic Mechanical Technology Co., Ltd., Fukushima, Japan) for 15 min.
142 The milled materials were passed through a 149- μ m sieve (Tokyo Screen Co. Ltd.,
143 Tokyo, Japan), and then milled APAP, TR-FB and intact MCC or milled MCC were

144 manually pre-mixed in a polyethylene bag for 5 min. Granulation was conducted
145 with a high-shear mixer (MECHANOMiLL, Okada Seiko Co., Ltd., Tokyo, Japan)
146 equipped with a rubber heater and temperature sensor as previously reported (Shiino
147 et al, 2012). The batch size was 130 g for all formulations; the formulation of each
148 batch is summarized in **Table 1**. To manufacture granules, the jacket temperature was
149 fixed at approximately 85 °C, and the IS was set to 1200 rpm. After the product
150 temperature reached 77 °C, the rubber heater was turned off, the lid was removed
151 and mixture was agitated at various IS for 1 min. To cool the obtained products, the
152 mixture was removed from the bed of the mixer, and spread into thin layers on metal
153 trays.

154

155 **2.5.2. Granules containing BRH**

156 Granules containing BRH were prepared by almost the same method as
157 APAP granules. Only two processes were different. One is that the size reduction of
158 BRH was performed using a different-type sample mill (SK-M, Kyoritsu-riko Co.,
159 Ltd., Tokyo, Japan) for 90 s. The other is that granulation was performed with
160 various IS as summarized in **Table 1**.

161

162 **2.6. Particle size distribution**

163 The particle size distribution of granules was evaluated by sieve analysis
164 using standard sieves (Tsutsui Scientific Instruments Co., Ltd., Tokyo, Japan) with
165 aperture sizes ranging from 54 to 1000 μm . The relative width of the particle size
166 distribution (R_w) was calculated by the following equation,

167
$$R_w = \frac{d_{90} - d_{10}}{d_{50}} \quad [1]$$

168 where d_{10} , d_{50} , and d_{90} are 10%, 50%, and 90% of the accumulated particle size under
169 a screen, respectively.

170

171 **2.7. Roundness**

172 One hundred granules with diameters in the range of 250–297 μm were
173 chosen randomly. Images of these granules were captured using an Olympus BHS
174 microscope (Olympus, Tokyo, Japan) connected to a digital imaging camera (Nikon
175 D80, Nikon, Tokyo, Japan). These images were analyzed with WinROOF image
176 analysis software (Version 5.5, Mitani, Fukui, Japan) to determine the exact
177 diameters and shapes of the granules. The shapes of the granules were defined by
178 their roundness (P_t/P_r), where P_t is the theoretical perimeter length of a perfectly

179 spherical granule with the same area as the one being analyzed, and P_r is the actual
180 perimeter length.

181

182 ***2.8. SEM observation of granules***

183 The surface morphology of MB particles and granules with a diameter range
184 of 250–297 μm was assessed by SEM as described above in section 2.4.

185

186 ***2.9. Drug release test***

187 The release behavior of APAP from granules with a diameter of 420–590
188 μm for each formulation was examined in accordance with the paddle method listed
189 in the Japanese Pharmacopoeia 16th Edition (JP 16th). The test medium was 900 mL
190 of distilled water. The medium temperature was set to 37.0 ± 0.5 °C and the paddle
191 speed was 100 rpm. At each time point, a 5-mL aliquot of the test solution was
192 withdrawn and replaced with an equal volume of buffer solution, and the aliquot was
193 passed through a membrane filter (Pore size: 0.45 μm , Toyo Roshi Kaisha Ltd.,
194 Tokyo, Japan). The amount of APAP released into the medium was quantitatively
195 determined by UV absorptiometry (UV-mini, Shimadzu, Kyoto, Japan) at 243 nm.

196 For BRH, granules with a diameter of 250–297 μm were tested. The test
197 medium was hydrochloride buffer solution (pH 1.2), and UV absorptiometry was
198 carried out at 245 nm. A series of dissolution tests for BRH granules was the same as
199 that for APAP granules.

200

201 ***2.10. Synchrotron X-ray computed tomography (CT) measurement***

202 BRH granules with diameters of 250–297 μm were put in Lindemann glass
203 capillaries with a diameter of 0.2–0.3 mm. X-ray CT measurements were performed
204 using a micro-CT instrument (Uesugi et al., 2012; Suzuki et al., 2011) installed at the
205 undulator beamline BL37XU of SPring-8 (Hyogo, Japan). The X-ray energy was set
206 to 8 keV, and flux density was approximately 6×10^{12} photons/s/mm². Nine hundred
207 transmission X-ray images, with parallel projection geometry, were recorded in 0.2°
208 steps with continuous rotation of the sample (on-the-fly scan mode). During
209 measurement, each sample was continuously irradiated with X-rays, and the
210 exposure time for each projection was 150 ms. The distance from the sample to the
211 area detector was 2 mm. CT measurement of one sample was accomplished within 5
212 min. All CT measurements were performed at room temperature. Tomographic
213 reconstruction was performed by the convolution-back-projection method using the

214 *CBP* package (Uesugi , 2004), and 1920×1920 pixel cross-sectional images were
215 obtained. One pixel is equivalent to $0.444 \times 0.444 \mu\text{m}$. 3-D analyses of the
216 cross-sectional images were performed using the *SLICE* package (Nakano et al.,
217 2006) and *ImageJ* (Schneider et al., 2012). X-ray linear attenuation coefficient
218 (LAC) values between 0 and 70 cm^{-1} are shown in 8-bit grayscale in the images of
219 granules, and LAC values higher than 70 cm^{-1} are in white.

220

221 ***2.11. Calculation of the void voxel ratio of granules***

222 To evaluate the porosity of the granules, the void voxel ratio (VVR) was
223 calculated from slice images of X-ray CT. VVR is defined as follows:

$$\text{VVR} = \frac{\text{(number of voxels regarded as void in the sample)}}{\text{(number of total voxels in the sample)}}$$

224 Voxels with LAC values less than 0.3 cm^{-1} were regarded as voids.

225

226 3. Results and discussion

227 3.1. Evaluation of TR-FB as an MB

228 3.1.1. Heat and storage stability of GM and TR-FB

229 First, to determine whether TR-FB works as an MB, the crystal states of TR-FB
230 and GM, which is generally used as an MB, were evaluated by DSC and PXRD.
231 **Figure 1** shows DSC curves of GM and TR-FB. In the first heating cycle, GM and
232 TR-FB exhibited endothermic peaks at 73.2 and 71.9 °C, respectively, and both
233 melted at a similar temperature. In the second heating cycle, the endothermic peak of
234 GM decreased to 67.5 °C because of the crystal transition from the stable β form to
235 the metastable α form, as reported by Yajima et al. (2002). GM also transforms from
236 the metastable form (α) to the stable form (β) through another metastable form (β')
237 during storage at ambient temperature (Windbergs et al., 2009). This crystal
238 transition is likely to change the drug dissolution profile of granules prepared with
239 GM after storage. In contrast, the endothermic peak of TR-FB did not shift in the
240 second heating cycle. The PXRD pattern of TR-FB was the same before and after
241 melting (**Fig. 2**). This indicates that TR-FB does not show any crystal transition in
242 the investigated temperature range. **Figure 3** shows SEM images of the surface of
243 GM and TR-FB particles prepared by a spray congealing method. After storage at

244 24 °C/25% RH for 6 weeks, acicular crystals were observed on the surface of GM
245 particles (**Fig. 3b**), consistent with previous observations (Windbergs et al., 2009).
246 Conversely, the TR-FB particle remained smooth even after storage (**Fig. 3d**). From
247 these results, TR-FB might be more stable as an MB than GM and should be suitable
248 for use in melt granulation.

249

250 *3. 1. 2. Effect of GM and TR-FB on granulation properties*

251 **Figure 4** shows the particle size distribution of granules prepared using GM
252 and TR-FB as an MB. In both types of granules, as the amount of GM or TR-FB
253 increased, the particle size of the granules also increased. **Table 2** shows the
254 physicochemical properties of the granules, such as d_{50} , R_w , yield of particles in the
255 106–210 μm range and roundness. The roundness of both types of granules was high
256 (over 0.83). When comparing the same amount of MB, d_{50} of TR-FB granules was
257 larger than that of GM granules; however, R_w of TR-FB granules was smaller, and
258 yield of particles in the 106–210 μm range of TR-FB is higher than that of GM
259 granules. This is because of the difference in the particle size and viscosity of melted
260 GM and TR-FB. The mean particle sizes of GM and TR-FB powders were 41 and 3.8
261 μm , respectively, while the viscosities of GM and TR-FB at 80 °C are 27.3 and 31.3

262 mPa·s, respectively. Previous literature suggested that a difference of viscosity of
263 about 4 mPa·s caused differences of physicochemical properties such as d_{50} and R_w
264 (Eliassen et al., 1999). The granule growth mechanism of melt granulation is shown in
265 **Fig. 5a**. Melted MB particles adhere to the powder, allowing the powder particles to
266 bond with each other to make core particles (distribution phase). Then, these core
267 particles adhere to each other (coalescence phase) and the granules become larger by
268 adhering repeatedly, suggesting that when the core particle is larger, the resulting
269 granules are larger. When the product is cooled and the temperature is close to the
270 melting point of the MB, the MB starts to congeal, and granules are sheared by the
271 impeller blade (**Fig. 5b**). Eliassen reported that when the viscosity of an MB is less
272 than 50 mPa·s, adhesion and breakage occur simultaneously during the coalescence
273 and agglomeration phase (Eliassen et al., 1999), so the particle size of granules
274 depends on the balance between adhesion and breakage. Therefore, in GM with a
275 large particle size, the core particles were large so the granules were also large.
276 Moreover, because of the low viscosity of GM, breakage frequently occurred in the
277 agglomeration phase to produce a large amount of finely crushed granules. As a
278 result, GM granules had a small d_{50} and large R_w . In contrast, for TR-FB with smaller
279 particle size and higher viscosity than those of GM, the core particles were smaller

280 and less breakage occurred, so the TR-FB granules had lower R_w and a larger yield of
281 fine granules with a particle size range of 106 to 210 μm than those of GM. **Figure 6**
282 shows drug dissolution profiles of GM and TR-FB. TR-FB granules showed more
283 sustained release than GM granules. Because the hydrophile-lipophile balance values
284 of GM and TR-FB are 4.0 and 2.1 respectively, TR-FB is more hydrophobic than
285 GM. Taken together, these findings suggest that TR-FB is a better MB than GM for
286 preparing fine granules with sustained drug release properties.

287

288 *3.2. Effect of MCC milling on granule properties*

289 *3.2.1. Particle size and external structure of granules*

290 **Figure 7** shows SEM images of intact and milled MCC particles. Intact
291 MCC particles were acicular, and their mean diameter was approximately 90 μm
292 (**Fig. 7a**). After milling, the MCC particles had an undefined shape and their mean
293 diameter was approximately 10 μm (**Fig. 7b**).

294 **Figure 8** shows the particle size distributions of granules prepared with
295 intact and milled MCC. **Table 3** shows physicochemical properties of the granules,
296 The granules containing 22.5% TR-FB and intact MCC (**Fig. 8a, Batch 7**) showed
297 similar particle size distribution to that of granules containing 17.5% TR-FB and

298 milled MCC (**Fig. 8b, Batch 12**). This indicates that even a small amount of TR-FB
299 was sufficient to prepare granules when milled MCC was used. For milled MCC,
300 addition of a larger amount of TR-FB and increasing IS increased granule size
301 compared with use of intact MCC, suggesting that careful control of operating
302 conditions is needed to prepare fine granules smaller than 200 μm . The roundness of
303 both types of granules was almost the same (**Table 3**). In both types of granules,
304 addition of a smaller amount of TR-FB and lower IS were found to make granules
305 rounder and *vice versa*. The granules prepared with either intact or milled MCC
306 using different binder concentration and IS had a similar appearance (**Fig. 9**).

307

308 **3. 2. 2. Drug dissolution from granules**

309 **Figure 10** shows dissolution profiles of BRH from granules containing
310 intact or milled MCC. For the milled MCC granules, it took about 30–60 min for
311 80% of the BRH to dissolve (**Fig. 10B**), while it took only about 15–30 min for 80%
312 of the BRH to dissolve from the intact MCC granules (**Fig. 10A**). Although the
313 appearance of these granules was almost the same (**Fig. 9**) and a larger amount of
314 TR-FB was present in the intact MCC granules than in the milled MCC ones, the
315 milled MCC granules exhibited more sustained drug release than the intact MCC

316 ones. This may be caused by differences in the internal structure of the granules, so
317 we conducted synchrotron X-ray CT measurements to obtain cross-sectional images
318 of the granules.

319

320 *3. 2. 3. Internal structure of granules*

321 **Figure 11** shows cross-sectional X-ray CT images of the granules. White
322 and black areas in each figure represent BRH (calculated LAC = 75.5 cm^{-1}) and void
323 space, respectively. Brighter gray and darker gray areas represent MCC (12.4 cm^{-1})
324 and TR-FB (4.1 cm^{-1}), respectively. TR-FB in intact MCC granules was present
325 mainly near the surface of the granules and adhered to MCC particles, which
326 produced voids inside the granules (**Fig. 11A-E**). Conversely, in the milled MCC-
327 granules, TR-FB surrounded milled MCC particles, and no large voids were
328 observed (**Fig. 11F-J**). This indicates that because intact MCC particles became core
329 particles and adhered to each other to form granules, voids were caused by the
330 acicular shape of intact MCC. In contrast, in milled MCC granules, because the
331 MCC particles were non-acicular and small, they were surrounded by TR-FB
332 efficiently, and granules with rigid structure were formed. Focusing on the
333 distribution of BRH, in intact MCC granules, BRH was found near the surface of the

334 granules, while BRH was uniformly distributed throughout milled MCC granules.

335 This is because the high hydrophobicity of BRH means that it has a high affinity for

336 hydrophobic TR-FB, indicating that the drug distribution in the granules depends on

337 the characteristics of the MB.

338 The VVR of each granule was then calculated (**Fig. 12**). In intact MCC

339 granules, VVR is significantly lower (26–27% to 15–17%) when the amount of MB

340 is higher (22.5% to 27.5%) ($p < 0.01$, compared with batch No. 7). Conversely, in

341 milled MCC granules, all of the granules exhibited markedly lower VVR (5–8%)

342 compared with that of the intact MCC granules (15–27%) ($p < 0.001$, compared with

343 batch No. 7). This difference of the porosity of the granules might cause the

344 difference of the drug release profiles of intact and milled MCC granules. **Figure 13**

345 shows the relationship between VVR and 80% drug release time from each granule.

346 Although these granules were prepared by different binder ratios, which affect the

347 drug dissolution ratio and by various impeller speeds while granulation, high

348 correlation coefficient value of liner regression was obtained. This is to say, because

349 of the higher porosity of intact MCC granules, test solution entered the granules

350 more readily, so the area where BRH was in contact with the test solution was larger

351 than that in the milled MCC granules. As a result, intact MCC granules released
352 BRH faster than milled MCC granules.

353 Up to now, it has been thought that melt granulation generally produces
354 granules with rigid structure because of the high speed of the impeller and high
355 centrifugation force regardless of formulation and operating conditions. However,
356 our results demonstrate that the milling process of the excipient affects the
357 distribution of drug particles and void space ratio in granules formed by high-shear
358 melt granulation, and these differences in the inner structure of granules affect drug
359 release.

360 **4. Conclusions**

361 We investigated TR-FB as an MB and the effect of milling MCC with the objective
362 of preparing fine granules with sustained drug release properties. TR-FB was more
363 stable during heating and storage than GM, a commonly used as MB. Granules
364 prepared with TR-FB were smaller, had a narrower particle size distribution, and
365 showed a better-sustained drug release profile compared with those formed with GM.
366 In addition, granules prepared with milled MCC had rigid structure, whereas those
367 produced with intact MCC had large void space. Consequently, the granules prepared
368 with milled MCC showed better-sustained drug release behavior than that of granules
369 formed with intact MCC, because penetration of test solution into granules was
370 limited. Our results reveal that TR-FB is suitable as an MB and milling MCC affects
371 the physicochemical properties and drug dissolution behavior of granules. Fine
372 granules with a diameter of less than 200 μm and sustained drug release behavior
373 were successfully prepared using TR-FB and milled MCC. In the near future, after
374 adding functional polymers to enhance the sustained release properties, further in
375 vivo studies would be necessary.

376

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446

447 **Table captions**

448 **Table 1.**

449 Formulations of granules and operating conditions.

450 **Table 2.**

451 Properties of particles prepared with GM (Batch No. 1-3) and TR-FB (Batch No.

452 4-6).

453 **Table 3.**

454 Properties of particles prepared with intact MCC (Batch No. 7-11) and milled MCC

455 (Batch No. 12-16).

456

457 **Table 1.**

Batch No.	Formulation						Operation
	APAP (%)	BRH (%)	GM (%)	TR-FB (%)	Intact MCC (%)	Milled MCC (%)	IS (rpm)
1	10	—	20.0	—	—	70.0	400
2	10	—	22.5	—	—	67.5	400
3	10	—	25.0	—	—	65.0	400
4	10	—	—	20.0	—	70.0	400
5	10	—	—	22.5	—	67.5	400
6	10	—	—	25.0	—	65.0	400
7	—	10	—	22.5	67.5	—	400
8	—	10	—	22.5	67.5	—	1200
9	—	10	—	27.5	62.5	—	400
10	—	10	—	27.5	62.5	—	1200
11	—	10	—	27.5	62.5	—	1600
12	—	10	—	22.5	—	67.5	400
13	—	10	—	22.5	—	67.5	1200
14	—	10	—	27.5	—	62.5	1600
15	—	10	—	27.5	—	62.5	400
16	—	10	—	27.5	—	62.5	1200

458

459

460 **Table 2.**

Batch No.	1	2	3	4	5	6
MB		GM			TR-FB	
Concentration (%)	20	22.5	25	20	22.5	25
d_{50} (μm)	101	105	195	125	160	290
R_w	4.16	3.41	2.91	1.92	1.78	2.27
Yield (%) (106-210 μm)	17.2	21.9	38.6	50.0	59.4	25.9
Roundness	0.86	0.89	0.88	0.86	0.86	0.83

461

462

463 **Table 3.**

Batch No.	7	8	9	10	11
Excipient	Intact MCC				
Binder (%)	22.5	22.5	27.5	27.5	27.5
IS (rpm)	400	1200	400	1200	1600
d_{50} (μm)	155	154	180	187	197
R_w	0.93	0.93	1.21	1.45	1.26
Yield (%) (106-210 μm)	66.7	66.9	56.9	51.5	47.0
Roundness	0.88	0.85	0.83	0.78	0.78

464

Batch No.	12	13	14	15	16
Excipient	Milled MCC				
Binder (%)	17.5	17.5	17.5	22.5	22.5
IS (rpm)	400	1200	1600	400	1200
d_{50} (μm)	149	197	185	351	421
R_w	1.28	1.90	2.25	1.90	1.53
Yield (%) (106-210 μm)	61.2	45.8	47.3	17.6	9.6
Roundness	0.89	0.81	0.83	0.80	0.79

465 **Figure legends**

466 **Fig. 1.** DSC curves of GM (**A, B**) and TR-FB (**C, D**) measured on first (**A, C**) and
467 second (**B, D**) heating cycles.

468 **Fig. 2.** PXRD patterns of TR-FB before (**A**) and after (**B**) heating.

469 **Fig. 3.** SEM images of the surface of MB particles prepared by a spray congealing
470 method using (**A, B**) GM and (**C, D**) TR-FB (**A, C**) before and (**B, D**) after storage
471 for 6 weeks.

472 **Fig. 4.** Effect of binder concentration on the particle size distributions of granules
473 prepared with (**A**) GM or (**B**) TR-FB.

474 **Fig. 5.** Diagram of the granulation mechanism in melt granulation (**A**) above and (**B**)
475 near the melting temperature of the MB.

476 **Fig. 6.** Drug dissolution profile from granules prepared with GM or TR-FB.

477 **Fig. 7.** SEM images of MCC (**A**) before and (**B**) after milling.

478 **Fig. 8.** Particle size distribution of granules prepared with (**A**) intact or (**B**) milled
479 MCC with various IS.

480 **Fig. 9.** SEM images of granules prepared with (**A-E**) intact or (**F-J**) milled MCC
481 with various IS.

482 **Fig. 10.** Drug dissolution profiles of granules prepared with (A) intact or (B) milled

483 MCC with various IS.

484 **Fig. 11.** X-ray CT images of granules prepared with (A-E) intact or (F-J) milled

485 MCC with various IS.

486 **Fig. 12.** VVR of granules prepared with intact or milled MCC with various IS. *:

487 $p < 0.01$ and **: $p < 0.001$, compared with VVR of batch No. 7.

488 **Fig. 13.** Relationship between VVR and 80% drug release time.